

**Figure 1 | A photonic analogue of a Floquet topological insulator.** Rechtsman *et al.*<sup>4</sup> have designed a hexagonal array of helical optical waveguides that functions as a photonic topological insulator. As an electromagnetic wave travels down the axes of the waveguides, it also moves sideways along the boundary of the array. This sideways motion is that of a ‘topological edge state’, and so is immune to back-scattering: on reaching the end of one edge of the array, the wave turns 90° and continues moving along the next edge. For simplicity, in this illustration the waveguides are wave-like rather than helical.

twist in the waveguides is formally equivalent to an oscillating potential, which, through Floquet’s theorem, yields topologically non-trivial bands. Their experiments directly demonstrate the existence of an electromagnetic wave that moves in a single direction around the edge of the waveguide array (Fig. 1), bypassing obstacles and imperfections exactly as predicted by the theory of topological insulators.

It is worth noting that this photonic topological insulator cannot be used as an optical isolator. The waveguide array is a

three-dimensional photonic crystal composed of non-magnetic material, and a well-known principle based on the time-reversal symmetry of such systems shows that they cannot act as isolators. Thus, for each edge wave travelling in one direction along the edge of the waveguide array, it is possible to excite another wave that moves backwards along the edge and backwards along the waveguide axis. (For those familiar with models of two-dimensional electronic topological insulators, the direction of propagation along the waveguide axis in this

system can be regarded as playing the part of the electron’s spin orientation.)

Although several other research groups have proposed different schemes for photonic topological insulators (see ref. 4 for references and ref. 12 for a recent experimental demonstration in an optical-chip platform), Rechtsman and colleagues’ method is notable for its simplicity and practicality. The robust properties of the topological edge waves indicate several possible device applications, such as carrying signals robustly through optical fibres. Future variants of this photonic topological insulator could also be used to explore many issues of fundamental scientific interest, including how the edge waves behave under conditions of nonlinearity, amplification and damping, all of which are easily achievable and tunable in photonic media. ■

**Yidong Chong** is in the School of Physical and Mathematical Sciences, Nanyang Technological University, Singapore 637371, Singapore.

e-mail: yidong@ntu.edu.sg

1. Yablonovitch, E. *Phys. Rev. Lett.* **58**, 2059–2062 (1987).
2. John, S. *Phys. Rev. Lett.* **58**, 2486–2489 (1987).
3. Joannopoulos, J. D., Johnson, S. G., Winn, J. N. & Meade, R. D. *Photonic Crystals: Molding the Flow of Light* (Princeton Univ. Press, 2008).
4. Rechtsman, M. C. *et al. Nature* **496**, 196–200 (2013).
5. Moore, J. E. *Nature* **464**, 194–198 (2010).
6. Bernevig, B. A. *et al. Science* **314**, 1757–1761 (2006).
7. König, M. *et al. Science* **318**, 766–770 (2007).
8. Wang, Z. *et al. Nature* **461**, 772–775 (2009).
9. Lindner, N. H. *et al. Nature Phys.* **7**, 490–495 (2011).
10. Fang, K. *et al. Nature Photon.* **6**, 782–787 (2012).
11. Schwartz, T. *et al. Nature* **446**, 52–55 (2007).
12. Hafezi, M., Fan, J., Migdall, A. & Taylor, J. Preprint at <http://arxiv.org/abs/1302.2153> (2013).

and Kim *et al.*<sup>3</sup> (page 219) address these questions using optogenetics to manipulate distinct neuronal subpopulations in mice and so dissect out the contribution of intermixed but functionally distinct cell groups\*.

Both teams analysed a large, diffuse brain region called the bed nucleus of the stria terminalis (BNST). Previous studies<sup>4–7</sup> have found that lesions of the BNST reduce anxiety and fear of specific environments. Other work has discovered<sup>8,9</sup> distinct subregions and subpopulations of BNST neurons, and has found that the region has connections with several other brain areas that are involved in motivated behaviour and stress responses. However, the functions of the various BNST subpopulations and subregions, as well as the significance of these connections, have remained unclear.

Jennings and colleagues focused on the role of the ventral BNST (vBNST) in mediating anxiety and regulating motivated behaviour, which, along with several other behaviours, may be modulated by anxiety. Consistent

\*This article and the papers under discussion<sup>2,3</sup> were published online on 20 March 2013.

## NEUROSCIENCE

# Anxiety is the sum of its parts

**Anxiety does not arise from a single neural circuit. An interplay between neighbouring, yet opposing, circuits produces anxiety, and outputs from these circuits regulate specific anxiety responses. SEE LETTERS P.219 & P.224**

JOSHUA P. JOHANSEN

**W**e all know anxiety. We might have experienced it while waiting to hear about a promotion at work, or on our way to see the doctor because she wants to talk about test results in person. A diffuse uneasiness, sometimes accompanied by perspiration and subtle changes in breathing, anxiety ebbs and flows depending on life’s circumstances, and can even occur for no

apparent reason. The condition can be healthy and adaptive, but research in the United States<sup>1</sup> shows that, for roughly one-third of people, anxiety is a debilitating disorder at some point in their lives. Nevertheless, answers to important questions — such as how different neuronal populations represent anxiety, and how the various components of the anxious state are constructed and represented in neural circuits — remain elusive. In two studies in this issue, Jennings *et al.*<sup>2</sup> (page 224)

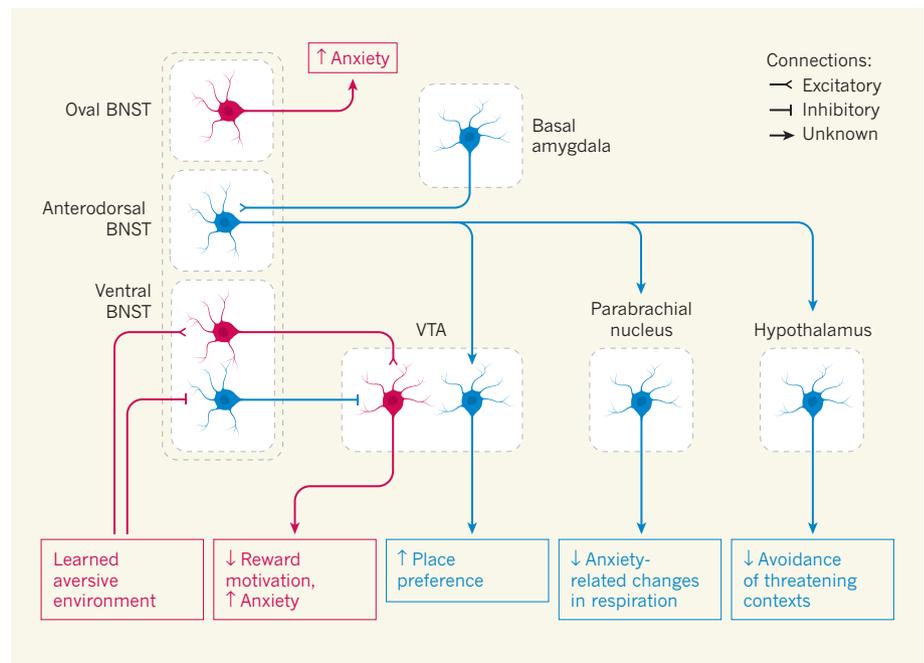
with the idea that the vBNST contains functionally distinct cell populations, the authors found that learned anxiety that is associated with specific environments leads to increased activity of some vBNST neurons and decreased activity of others.

Both of these cell populations made specific synaptic connections with neurons of another brain region called the ventral tegmental area (VTA), which is known to guide motivated behaviour. Specifically, cells that were excited by anxiety-inducing environments in turn excited their VTA partner, and stimulating these excitatory vBNST–VTA connections increased anxiety and decreased reward-seeking behaviour. By contrast, vBNST neurons that were inhibited by anxiety-inducing environments also inhibited their downstream VTA neurons, and stimulating these inhibitory connections promoted reward-seeking behaviour and reduced anxiety.

A caveat of this work is that the authors did not inhibit vBNST–VTA connections during natural anxiety states, but rather stimulated the neurons to regulate anxiety and motivated behaviours. Thus, it is possible that engagement of these circuits by anxiety does not produce the same behavioural effects naturally as those seen with artificial stimulation. However, the fact that during anxious states the vBNST–VTA neurons, which are known to promote anxiety, were activated and those that reduce anxiety were inhibited provides strong correlative evidence that learned anxiety naturally engages these neuronal subpopulations. The interplay between these two opposing ‘push–pull’ circuits may set an adaptive, or even a maladaptive, level of anxiety, and allow for bidirectional regulation of reward-motivated behaviour during anxiety.

Kim and co-workers asked whether, and if so how, cells in the two subregions of the dorsal BNST, the oval nucleus (ovBNST) and the anterodorsal BNST (adBNST), differentially regulate anxiety. They found that the activity of ovBNST neurons promoted anxiety. Moreover, inputs from the amygdala, a brain region that has been implicated in fear, reward and anxiety, activated adBNST neurons and reduced anxiety, and inhibition of these inputs increased anxiety. Consistent with a role in reducing anxiety, adBNST neurons fired more when the animals were in a safe environment than when they were in an anxiety-producing one, thus distinguishing between the two places (Fig. 1).

Intriguingly, inhibiting amygdala inputs to the adBNST reduced the ability of this subregion’s neurons to distinguish between safe and anxiety-producing places, which suggests that adBNST cells reduce anxiety in response to a ‘safety’ signal from the amygdala. Future work should determine how amygdala neurons connecting to the adBNST encode anxiety-related information and what types of experience recruit this anxiety-reducing circuit.



**Figure 1 | Multiple personalities of an anxiety circuit.** Two studies<sup>2,3</sup> show that various subregions in the bed nucleus of the stria terminalis (BNST) of the mouse brain contain intermixed cell populations that can produce (red) or ameliorate (blue) anxiety in a modular manner. Within the dorsal BNST, outputs from the oval nucleus promote anxiety, whereas outputs from the anterodorsal BNST — driven by activity in the amygdala — reduce anxiety. Anterodorsal BNST neurons also make specific connections to other brain regions, such as the hypothalamus, parabrachial nucleus and ventral tegmental area (VTA), to ameliorate specific features of anxiety. The ventral BNST contains intermixed but functionally distinct subpopulations of neurons. When the mice are exposed to a known anxiety-causing environment, some of these neurons are excited, stimulating their connections in the VTA to produce anxiety and reduce reward motivation. Other ventral BNST neurons that reduce anxiety and enhance reward motivation are inhibited, facilitating the production of anxiety.

Kim *et al.* also examined specific connections between the adBNST and other brain regions and found that, depending on the connections involved, the adBNST reduced specific aspects of the anxiety response. For instance, stimulating the connections between the adBNST and the hypothalamus reduced the tendency of mice to avoid anxiety-producing places; stimulating connections to neurons of the parabrachial nucleus led to reduced anxiety-induced changes in respiration; and stimulating connections with VTA neurons resulted in place preference (Fig. 1).

The two studies give us a richer understanding of how anxiety is represented by opposing but complementary neural circuits in the BNST. They highlight the modular nature of anxiety circuits and suggest a concerted mechanism for bidirectional regulation of anxiety-related responses. This type of bidirectional coding has been seen in other parts of the anxiety circuit<sup>10,11</sup>, particularly in the brain’s medial prefrontal cortex, in which single neurons differentially represent safe and anxiety-producing environments.

In fact, this type of circuit design may be a general feature of both fear and anxiety systems. There is strong evidence<sup>12,13</sup> that partially distinct neuronal subpopulations mediate fear and safety-from-fear learning. Moreover, fear and anxiety are closely related

conceptually, and brain regions such as the amygdala, medial prefrontal cortex, hippocampus and BNST are involved in both. Understanding the principles shared by the two systems, and how their respective neural circuits interact, will be research areas of great interest for the future. ■

**Joshua P. Johansen** is at the *RIKEN Brain Science Institute, Saitama 351-0198, Japan.*  
e-mail: [jjohans@brain.riken.jp](mailto:jjohans@brain.riken.jp)

- Kessler, R. C. *et al. Arch. Gen. Psychiatry* **62**, 593–602 (2005).
- Jennings, J. H. *et al.* **496**, 224–228 (2013).
- Kim, S.-Y. *et al. Nature* **496**, 219–223 (2013).
- Davis, M., Walker, D. L., Miles, L. & Grillon, C. *Neuropsychopharmacology* **35**, 105–135 (2010).
- Sullivan, G. M. *et al. Neuroscience* **128**, 7–14 (2004).
- Duvarci, S., Bauer, E. P. & Pare, D. *J. Neurosci.* **29**, 10357–10361 (2009).
- Poulos, A. M., Ponnusamy, R., Dong, H. W. & Fanselow, M. S. *Proc. Natl Acad. Sci. USA* **107**, 14881–14886 (2010).
- Kudo, T. *et al. J. Neurosci.* **32**, 18035–18046 (2012).
- Dong, H.-W. & Swanson, L. W. *J. Comp. Neurol.* **468**, 277–298 (2004).
- Adhikari, A., Topiwala, M. A. & Gordon, J. A. *Neuron* **71**, 898–910 (2011).
- Tye, K. M. *et al. Nature* **471**, 358–362 (2011).
- Maren, S. & Quirk, G. J. *Nature Rev. Neurosci.* **5**, 844–852 (2004).
- Herry, C. *et al. Eur. J. Neurosci.* **31**, 599–612 (2010).